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(54) Title: MULTIPLE VIRUS REPLICON CULTURE SYSTEMS

(57) Abstract: Methods and compositions are provided for screening candidate antiviral agents using cells containing subgenomic viral replication systems such as replicons and minigenomes. The methods involve the simultaneous assay of more than one subgenomic viral replication system. Compositions useful for theses methods are also provided.

- 9. Burgart, L. J., R. A. Robinson, M. J. Heller, W. W. Wilke, O. K. Iakoubova, and J. C. Cheville 1992. Multiplex polymerase chain reaction Mod Pathol. 5:320-3.
- Camenisch, G., M. Gruber, G. Donoho, P. Van Sloun, R. H. Wenger, and M. Gassmann 1996. A polyoma-based episomal vector efficiently expresses exogenous genes in mouse embryonic stem cells Nucleic Acids Res. 24:3707-13.
 - 11. Clarke, D. K., M. S. Sidhu, J. E. Johnson, and S. A. Udem 2000. Rescue of mumps virus from cDNA J Virol. 74:4831-8.
- 12. Collins, P. L., M. G. Hill, E. Camargo, H. Grosfeld, R. M. Chanock, and B. R. Murphy 1995. Production of infectious human respiratory syncytial virus from cloned cDNA confirms an essential role for the transcription elongation factor from the 5' proximal open reading frame of the M2 mRNA in gene expression and provides a capability for vaccine development Proc Natl Acad Sci U S A. 92:11563-7.
 - 13. Collins, P. L., M. G. Hill, J. Cristina, and H. Grosfeld 1996. Transcription elongation factor of respiratory syncytial virus, a nonsegmented negative-strand RNA virus Proc. Natl. Acad. Sci. U S A. 93:81-85.
- 14. Constantinescu, N. 1991. [Molecular mechanisms of the antiviral effect of interferon] Rev Roum Virol. 42:191-213.
- Conzelmann, K. K., and M. Schnell 1994. Rescue of synthetic genomic RNA analogs of rabies virus by plasmid- encoded proteins J Virol. 68:713-9.
 - 16. Corey, D. R. 1997. Peptide nucleic acids: expanding the scope of nucleic acid recognition Trends Biotechnol. 15:224-9.
- Davis, N. L., F. B. Grieder, J. F. Smith, G. F. Greenwald, M. L. Valenski, D. C. Sellon, P. C. Charles, and R. E. Johnston 1994. A molecular genetic approach to the study of Venezuelan equine encephalitis virus pathogenesis Arch Virol Suppl. 9:99-109.
- Davis, N. L., L. V. Willis, J. F. Smith, and R. E. Johnston 1989. In vitro synthesis of infectious venezuelan equine encephalitis virus RNA from a cDNA clone: analysis of a viable deletion mutant Virology. 171:189-204.
- dos Santos, C. N., P. R. Post, R. Carvalho, Ferreira, II, C. M. Rice, and R. Galler 1995. Complete nucleotide sequence of yellow fever virus vaccine strains 17DD and 17D-213 Virus Res. 35:35-41.
- Dryga, S. A., O. A. Dryga, and S. Schlesinger 1997. Identification of mutations in a Sindbis virus variant able to establish persistent infection in BHK cells: the importance of a mutation in the nsP2 gene Virology. 228:74-83.
 - 21. Elnifro, E. M., A. M. Ashshi, R. J. Cooper, and P. E. Klapper 2000. Multiplex PCR: optimization and application in diagnostic virology Clin Microbiol Rev. 13:559-70.

35

- 34. Kapoor, M., L. Zhang, P. M. Mohan, and R. Padmanabhan 1995. Synthesis and characterization of an infectious dengue virus type-2 RNA genome (New Guinea C strain) Gene. 162:175-80.
- Khromykh, A. A., and E. G. Westaway 1997. Subgenomic replicons of the flavivirus Kunjin: construction and applications J Virol. 71:1497-505.
- Kinney, R. M., S. Butrapet, G. J. Chang, K. R. Tsuchiya, J. T. Roehrig, N.
 Bhamarapravati, and D. J. Gubler 1997. Construction of infectious cDNA clones for dengue 2 virus: strain 16681 and its attenuated vaccine derivative, strain PDK-53 Virology. 230:300-8.
- Kwong, A. D., and N. Frenkel 1984. Herpes simplex virus amplicon: effect of size on replication of constructed defective genomes containing eucaryotic DNA sequences J Virol. 51:595-603.
- Lai, C. J., B. T. Zhao, H. Hori, and M. Bray 1991. Infectious RNA transcribed from stably cloned full-length cDNA of dengue type 4 virus Proc Natl Acad Sci U S A.
 88:5139-43.
- Lee, K. J., I. S. Novella, M. N. Teng, M. B. Oldstone, and J. C. de La Torre 2000. NP and L proteins of lymphocytic choriomeningitis virus (LCMV) are sufficient for efficient transcription and replication of LCMV genomic RNA analogs J Virol. 74:3470-7.
 - 40. Liljestrom, P., and H. Garoff 1991. A new generation of animal cell expression vectors based on the Semliki Forest virus replicon Biotechnology (N Y). 9:1356-61.
- Lindberg, A. M., C. Polacek, and S. Johansson 1997. Amplification and cloning of complete enterovirus genomes by long distance PCR J Virol Methods. 65:191-9.
 - 42. Lindenbach, B. D., and C. M. Rice 1997. trans-Complementation of yellow fever virus NS1 reveals a role in early RNA replication J Virol. 71:9608-17.
 - 43. Lohmann, V., F. Korner, A. Dobierzewska, and R. Bartenschlager 2001. Mutations in hepatitis C virus RNAs conferring cell culture adaptation J Virol. 75:1437-49.
- 44. Lohmann, V., F. Korner, J. Koch, U. Herian, L. Theilmann, and R. Bartenschlager
 1999. Replication of subgenomic hepatitis C virus RNAs in a hepatoma cell line [see comments] Science. 285:110-3.
- Lu, B., and H. J. Federoff 1995. Herpes simplex virus type 1 amplicon vectors with glucocorticoid- inducible gene expression Hum Gene Ther. 6:419-28.
 - 46. MacDonald, G. H., and R. E. Johnston 2000. Role of dendritic cell targeting in Venezuelan equine encephalitis virus pathogenesis J Virol. 74:914-22.

45

- 59. Podevin, P., A. Sabile, R. Gajardo, N. Delhem, A. Abadie, P. Y. Lozach, L. Beretta, and C. Brechot 2001. Expression of hepatitis C virus NS5A natural mutants in a hepatocytic cell line inhibits the antiviral effect of interferon in a PKR- independent manner Hepatology. 33:1503-11.
 - 60. Polo, S., G. Ketner, R. Levis, and B. Falgout 1997. Infectious RNA transcripts from full-length dengue virus type 2 cDNA clones made in yeast J Virol. 71:5366-74.
- Pugachev, K. V., M. S. Galinski, and T. K. Frey 2000. Infectious cDNA clone of the RA27/3 vaccine strain of Rubella virus Virology. 273:189-97.
 - 62. Pur, B., S. Polo, C. G. Hayes, and B. Falgout 2000. Construction of a full length infectious clone for dengue-1 virus Western Pacific, 74 strain Virus Genes. 20:57-63.
- Racaniello, V. R. 1984. Studying poliovirus with infectious cloned cDNA Rev Infect Dis. 6 Suppl 2:S514-5.
- Randhawa, J. S., A. C. Marriott, C. R. Pringle, and A. J. Easton 1997. Rescue of synthetic minireplicons establishes the absence of the NS1 and NS2 genes from avian pneumovirus J Virol. 71:9849-54.
- Rice, C. M., A. Grakoui, R. Galler, and T. J. Chambers 1989. Transcription of infectious yellow fever RNA from full-length cDNA templates produced by in vitro ligation New Biol. 1:285-96.
- Rice, C. M., R. Levis, J. H. Strauss, and H. V. Huang 1987. Production of infectious RNA transcripts from Sindbis virus cDNA clones: mapping of lethal mutations, rescue of a temperature-sensitive marker, and in vitro mutagenesis to generate defined mutants J Virol. 61:3809-19.
 - Roner, M. R., and W. K. Joklik 2001. Reovirus reverse genetics: Incorporation of the CAT gene into the reovirus genome Proc Natl Acad Sci U S A. 98:8036-41.
- Ryman, K. D., W. B. Klimstra, K. B. Nguyen, C. A. Biron, and R. E. Johnston 2000. Alpha/beta interferon protects adult mice from fatal Sindbis virus infection and is an important determinant of cell and tissue tropism J Virol. 74:3366-78.
- Saito, S. 1990. Enhancement of the interferon-induced double-stranded RNA-dependent protein kinase activity by Sindbis virus infection and heat-shock stress Microbiol Immunol. 34:859-70.
 - 70. Schlesinger, S., and T. W. Dubensky 1999. Alphavirus vectors for gene expression and vaccines Curr Opin Biotechnol. 10:434-9.
- 71. Schranz, P., H. Zentgraf, and C. H. Schroder 1990. Integrated defective replication units of hepatitis B virus Virus Genes. 4:367-74.

30

35

nonstructural protein NSs inhibits viral RNA synthesis in a minireplicon system Virology. 281:67-74.

- Weiss, L., A. S. Kekule, U. Jakubowski, E. Burgelt, and P. H. Hofschneider 1996.

 The HBV-producing cell line HepG2-4A5: a new in vitro system for studying the regulation of HBV replication and for screening anti- hepatitis B virus drugs Virology. 216:214-8.
- Whelan, S. P., L. A. Ball, J. N. Barr, and G. T. Wertz 1995. Efficient recovery of infectious vesicular stomatitis virus entirely from cDNA clones Proc Natl Acad Sci U S A. 92:8388-92.
- Widell, A., B. G. Hansson, B. Oberg, and E. Nordenfelt 1986. Influence of twenty potentially antiviral substances on in vitro multiplication of hepatitis A virus Antiviral Res. 6:103-12.
- Wychowski, C., S. U. Emerson, J. Silver, and S. M. Feinstone 1990. Construction of recombinant DNA molecules by the use of a single stranded DNA generated by the polymerase chain reaction: its application to chimeric hepatitis A virus/poliovirus subgenomic cDNA Nucleic Acids Res. 18:913-8.
 - 89. Zhang, F., Q. Huang, W. Ma, S. Jiang, Y. Fan, and H. Zhang 2001. Amplification and cloning of the full-length genome of Japanese encephalitis virus by a novel long RT-PCR protocol in a cosmid vector J Virol Methods. 96:171-82.

Primary screening programs to discover and identify compounds with antiviral activity can be designed in a variety of ways. All programs, however, fall into one of two general approaches. In the targeted approach, one particular biochemical target is chosen and candidate antiviral compounds are screened for inhibition of that target. The target is often an enzyme or a receptor that is known or thought to be essential to the process of viral replication. The alternative approach is unbiased such that inhibitors of viral replication are sought without a priori concern for the target. This unbiased approach generally involves use of cell culture since, as obligate intracellular pathogens, viruses can only replicate within cells. Although cell-based screening has been used successfully throughout the drug-discovery field, it is problematic when screening for antivirals. This is because it requires inoculation of infectious virus onto the cells and the production of additional infectious progeny virus. In particular, handling such infectious material is not easily compatible with the high throughput process of screening large libraries of compounds.

Thus, there is a need for improved methods and compositions that are useful for screening and analyzing antiviral compounds. In particular, these methods and compositions should be useful for high-throughput antiviral screening. The invention described herein satisfies